

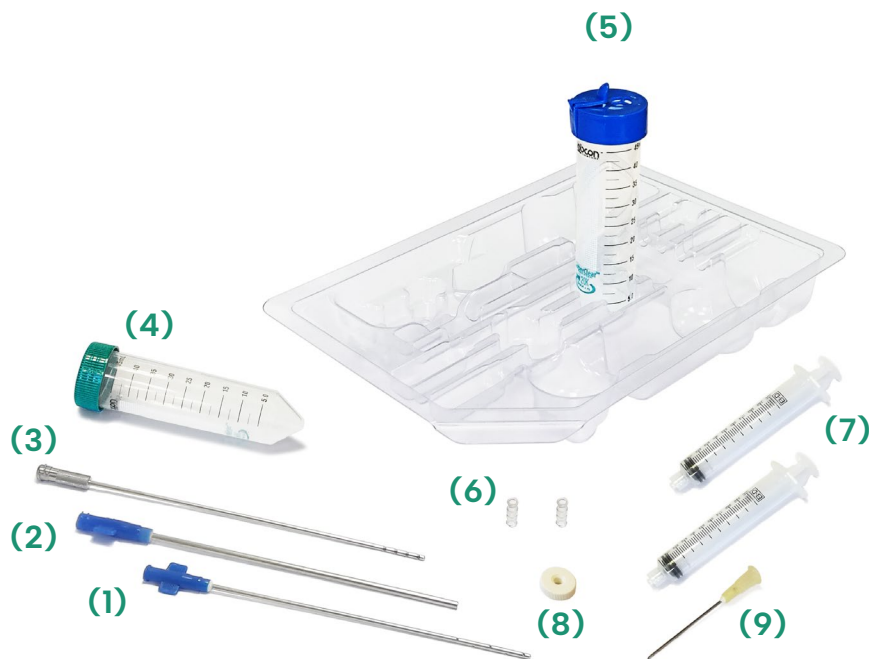


**All-in-one, sterile and single use  
fat transfer kits**

Procedure for the Myfill kit

Stemcis

## Myfill kit<sup>1</sup>



### Instruments

- (1) 1 infiltration cannula  
10 holes (Ø 1.0 mm)  
(Ø 2.1 mm x L150 mm)
- (2) 1 transfer cannula  
(Ø 3.0 mm x L130 mm)
- (3) 1 harvesting cannula  
8 holes (Ø 1.2mm)  
(Ø 1.7 - 2.1 mm x L150 mm)
- (4) 1 balance tube – 50 mL
- (5) 1 centrifugation tube – 50 mL
- (6) 2 FLL-FLL connectors
- (7) 6 Luer-Lock syringes – 10 mL  
with removable plunger
- (8) 1 rotor nut
- (9) 1 needle  
19G

### Additional products

We recommend gentle, short centrifugation for better survival of the fat graft.

Our centrifuge is compatible with all Stemcis kits thanks to different rotors.



### Adip'Spin Pre-programmed centrifuge

To change the programme, press the OPEN button for 8 sec.



### Rotor R6

Rotor R6, with the adaptators, compatible with the Myfill.

### PREPARATION

#### Requirements

- Stemcis centrifuge (Adip'Spin) with rotor R6.
- Two sterile bowls (one bowl containing the 0.9% NaCl and the other bowl is required for the waste NaCl material.)
- Approximately 200 mL of 0.9% NaCl are required for the "washing" step.
- One 500 mL bag (sterile) of injectable 0.9% NaCl for infiltration – **see recommended composition below** (one extra bowl may be required if you wish to have the NaCl in a bowl)
- One scalpel blade (#11), one sterile field and one Myfill kit

#### Getting started

- Switch the Adip'Spin centrifuge on (the switch is located at the rear).
- Check that the centrifuge is on the «FAT» programme.
- Open the Myfill kit
- Install the pre-sterilised rotor to the Adip'Spin centrifuge. Secure it with the white nut (supplied).
- Insert two metal sleeves vertically (opposing sides), then rotate the top of the sleeve towards the centre (this will allow the sleeve to slide down).



### INFILTRATION

#### Recommended composition of the infiltration liquid

- 500 mL of 0.9% NaCl
- 1 mg of adrenaline
- (20 mL of 2% lidocaine, only in case of local anaesthesia)

A ratio ranging between **1:1 to 2:1** is recommended (infiltrate 100 mL– 200 mL per 100 mL of sampled tissue).

Puncture the skin with the scalpel blade and infiltrate the harvesting area using the infiltration cannula and an "IV giving set" or a 10 mL syringe.



Harvesting can commence 5 - 10 minutes after infiltrating the harvesting area.

### HARVESTING

- Perform liposuction using the harvesting cannula connected to a 10 mL syringe and harvest the tissue at **2 mL increments**.



Never exceed a vacuum greater than 2 mL in the syringe. The negative pressure will result in damaged cells, indicative by oil in the syringe after decanting.

**NOTE: At the beginning, a few cycles are necessary to remove the infiltration solution. Dispose of the solution in the waste bowl.**

- Leave the syringe full of adipose tissue to decant for **one or two minutes** in the plastic tray.

## Procedure for the Myfill kit

### HARVESTING

- When two layers are clearly visible in the syringe, **drain** the bottom layer by pushing the plunger. **Retain the adipose tissue** (top layer) in the syringe.

**NOTE: Syringe must be kept upright, do not mix contents.**

- Place the T tube (blue tube) on the plastic tray and open the lid. Attach the transfer cannula to the syringe and fill to a maximum of 35 mL per tube.

### WASHING & CENTRIFUGATION

- Fill the T tube to the 45 mL mark (irrespective of the amount of adipose tissue in the tube) using a 10 mL syringe connected to the transfer cannula and containing 0.9% NaCl. Close the lids of the T tube.

**NOTE: Only the T tube (blue) contains adipose tissue; fill the second tube (green) to the 45 mL mark to ensure the centrifuge is balanced.**

- Gently invert / rotate the T tube 3 to 5 times to mix the contents.
- Place the tubes in the adaptors in the Adip'Spin centrifuge, the blue lid towards the outside. The rotor must be **correctly balanced**. All of the tubes must be filled to 45 mL.
- Start the centrifugation by pressing **"RUN"**. The first rectangle shows solid black at the end of the first centrifugation.
- Remove the T tube from the centrifuge. Keep it upright and place it on the plastic tray. Open the lid and connect a 10 mL syringe to the transfer cannula. Gently penetrate the adipose tissue until you reach the fluids at the bottom. Remove the fluids (only adipose tissue must remain in the tube). Empty the syringe.

**Repeat WASH steps twice.**

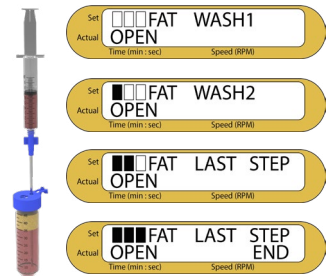
**NOTE: If you only require two washes (no local anaesthetic), then you can skip the second wash setting: therefore, perform two successive centrifugations (wash two and three) without removing the tubes. If tissue is bloody then three washes are required.**

**The last wash "last step" is very important – Do not skip this wash**

**NOTE: In the case of oil (after the last wash), remove the oil with the transfer cannula connected to a 10 mL syringe (before removing the fluids).**

- Extract the adipose tissue with the transfer cannula connected to a 10 mL syringe.

**Avoid the injection of oil.**



### INJECTION

- Inject the adipose tissue using the **19G needle** (supplied).





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